

Bio 127 - Section II
Early Development and Cell Fate
Determination

- I. Fertilization and Cleavage
- II. Specification and Gastrulation
- III. Organizing Power and Axis Formation

A. Some Really Big Ideas....

- 1. Species Specificity Must be Maintained
- 2. A Single Sperm is all You Need (or Want)
- 3. Genetic Fusion Signals Major Changes
- 4. The Egg is Built for Early Development

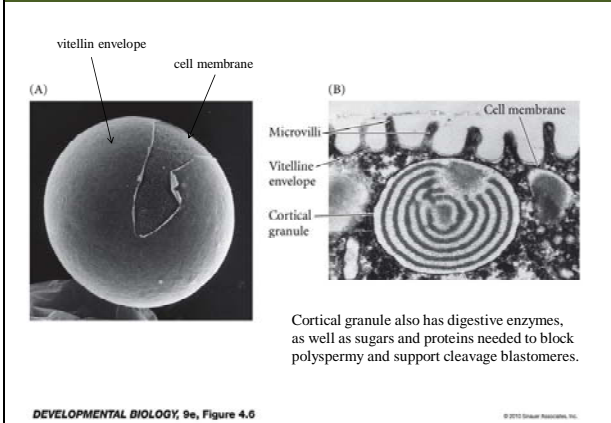
B. The Structures of the Gametes

- The sperm is built for species identification and high speed DNA delivery
- The egg is built to receive only one sperm and to store everything needed to get started down the path to development

Remember, the egg is complexly organized....

- The sea urchin egg is 10,000 times the volume of the sperm (ostrich eggs – wow!)
 - Nutrition
 - mRNA
 - Transcription factors
 - Ribosomes and tRNA
 - Secretable paracrine factors
 - Protection: DNA repair, distaste, antibodies

Structures at the plasma membrane



C. External Fertilization is Better Understood

- Even today much of what we truly understand about fertilization has come from the study of organisms who fertilize their eggs outside of the female's body
- Understanding internal fertilization is big-money big-science

Activation of egg metabolism occurs in the cytosol, independent of the pronuclei

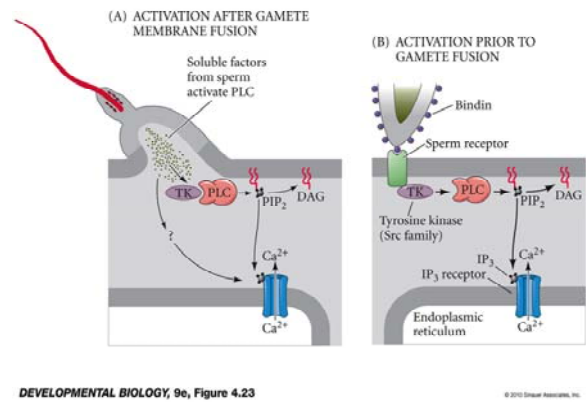


Figure 4.26 Postulated pathway of egg activation in the sea urchin (Part 2)

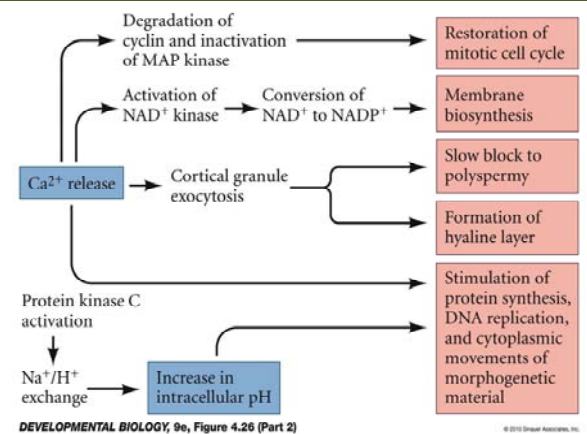


TABLE 4.1 Events of sea urchin fertilization (Part 1)

Event	Approximate time postinsemination ^a
EARLY RESPONSES	
Sperm-egg binding	0 seconds
Fertilization potential rise (fast block to polyspermy)	within 1 sec
Sperm-egg membrane fusion	within 1 sec
Calcium increase first detected	10 sec
Cortical granule exocytosis (slow block to polyspermy)	15–60 sec

Main sources: Whitaker and Steinhardt 1985; Mohri et al. 1995.
^aApproximate times based on data from *S. purpuratus* (15–17°C), *L. pictus* (16–18°C), *A. punctulata* (18–20°C), and *L. variegatus* (22–24°C). The timing of events within the first minute is best known for *Lytechinus variegatus*, so times are listed for that species.

TABLE 4.1 Events of sea urchin fertilization (Part 2)

Event	Approximate time postinsemination*
LATE RESPONSES	
Activation of NAD kinase	starts at 1 min
Increase in NADP ⁺ and NADPH	starts at 1 min
Increase in O ₂ consumption	starts at 1 min
Sperm entry	1–2 min
Acid efflux	1–5 min
Increase in pH (remains high)	1–5 min
Sperm chromatin decondensation	2–12 min
Sperm nucleus migration to egg center	2–12 min
Egg nucleus migration to sperm nucleus	5–10 min
Activation of protein synthesis	starts at 5–10 min
Activation of amino acid transport	starts at 5–10 min
Initiation of DNA synthesis	20–40 min
Mitosis	60–80 min
First cleavage	85–95 min

Main sources: Whitaker and Steinhardt 1988; Mohri et al. 1995.

*Approximate times based on data from *S. purpuratus* (15–17°C), *L. pictus* (16–18°C), *A. punctulata* (18–20°C), and *L. variegatus* (22–24°C). The timing of events within the first minute is best known for *Lytechinus variegatus*, so times are listed for that species.

DEVELOPMENTAL BIOLOGY, 9e, Table 4.1 (Part 2)

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Fusion of the Genetic Material in Sea Urchins

- Sperm carries its nucleus, centriole, mitochondria and flagellum into egg
 - Mito's and flagellum disintegrate in cytosol
 - Interestingly, in mice, centriole also degrades
- DNA decondenses to form pronucleus
 - First the membrane breaks up, exposing DNA
 - Then sperm's histones are replaced by egg's
 - This loosens up nucleosomes for replication

Fusion of the Genetic Material in Sea Urchins

- Male pronucleus then aligns with its centriole between it and female pronucleus
- The centriole sends out microtubules that integrate with egg's to form an aster between the two pronuclei
- The pronuclei are then pulled together to allow fusion and formation of the diploid zygote nucleus

D. What We Know of Internal Fertilization

- Tough to study
 - Fertilization occurs in the female oviducts
 - We don't yet know all conditions sperm encounter
 - Sperm is ejaculated at nearly every developmental stage
 - Of 280 million, 200 reach the egg
 - We don't yet know why the winners are the winners

The Female Reproductive Tract Aids Transport and Maturity of Gametes

- The ampulla is the region of oviduct where fertilization takes place
 - Uterine contractions help get sperm there
 - There is a holding region just before ampulla
 - Sperm flagella are stimulated near egg
 - Directional cues coming from egg or cummulus
 - Along the migration route sperm are stimulated to mature "Capacitation"

Figure 4.29 Hypothetical model for mammalian sperm capacitation

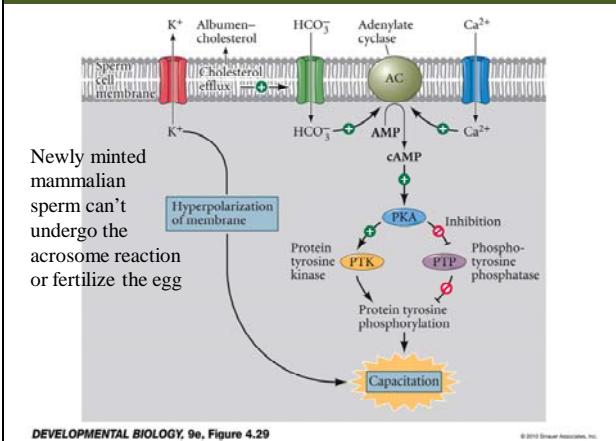
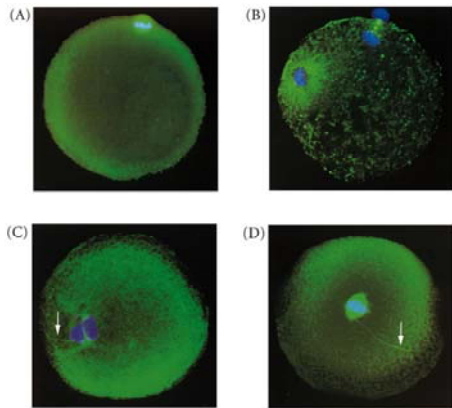


Figure 4.35 Pronuclear movements during human fertilization



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II. Cleavage

A. Background Information

B. Invertebrates

1. Sea Urchins
2. Snails
3. Tunicates
4. C. Elegans
5. Drosophila melanogaster

C. Vertebrates

1. The Frog
2. Zebrafish
3. The Chick Embryo
4. Mammals

A. Background Information

1. The Model Organisms
2. Structure-Process-Structure
3. Rapid Mitotic Cell Divisions
4. Cleavage Patterns
5. Mid-Blastula Transition

1. The Model Organisms

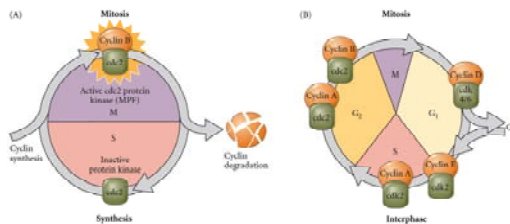
- a. Echinoderms (sea urchins)
- b. Gastropod molluscs (snails)
- c. Tunicates (ascidians)
- d. Nematode worms (*C. elegans*)
- e. Insects (*D. melanogaster*)
- f. Amphibians (*Xenopus laevis*)
- g. Fish (*Danio rerio*)
- h. Avians (*Gallus gallus*)
- i. Mammals (human and mouse)

Cleavage is a developmental process that takes the organism from fertilization through the blastula stage

Structure → Process → Structure

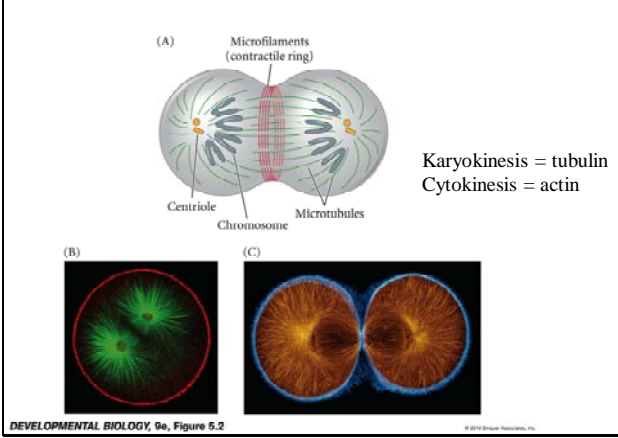
Cleavage is rapid mitotic cell division

The G-phases of somatic mitosis allow for cell growth so that the daughter cells are equal in size to the parent cell. In blastomeres we are trying to reduce the volume of the egg to somatic levels.



Frogs can make 37,000 cells in 43 hours.
Fruit flies can make 50,000 in 12 hours (10 min!)

Figure 5.2. Role of microtubules and microfilaments in cell division



Mid-Blastula Transition

G-phases added back as cleavage goes on.

- Xenopus adds G1 and G2 back after 12 round
- Drosophila adds G1 at round 14 and G2 at 17

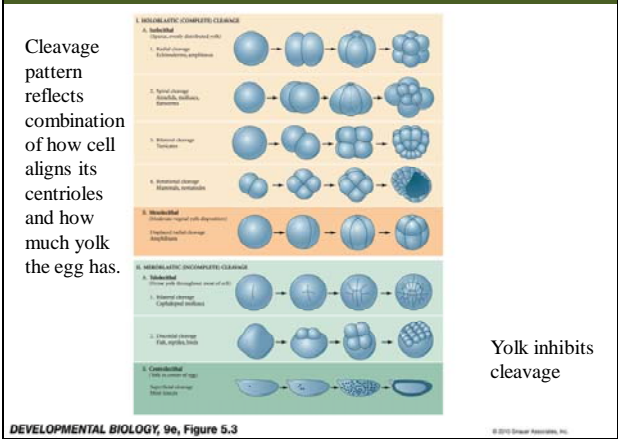
Synchronicity is lost as cells “go own way”

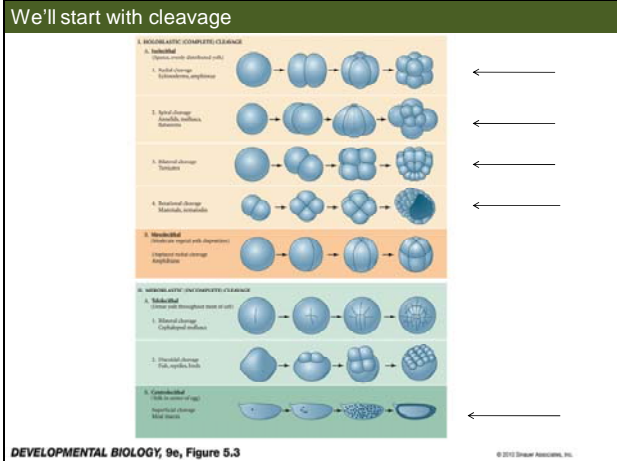
- Different regions of egg produce different cycle controls

New mRNAs are transcribed

- The beginnings of true differentiation

Figure 5.3 Summary of the main patterns of cleavage

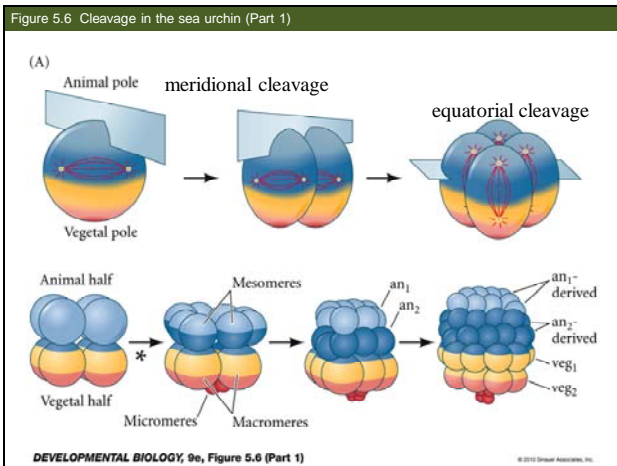




B. Invertebrates

1. We'll start with the sea urchin....

- Holoblastic: all of the egg forms into cells
- Isolecithal: sparse yolk throughout cytosol
- Animal-Vegetal: a little more yolk in vegetal end



- C1: meridional and C2: meridional
- C3: equatorial
- C4: animal pole 4 divide meridionally
 - vegetal 4 divide unequally equatorial
- C5: animal 8 divide equatorially
 - vegetal 4 macros divide meridionally
 - vegetal 4 micros divide unequally
- C6: animal 16 divide meridionally
 - vegetal divide equatorially
- C7: animal 16 divide equatorially
 - vegetal divide meridionally

Figure 5.6 Cleavage in the sea urchin (Part 2)

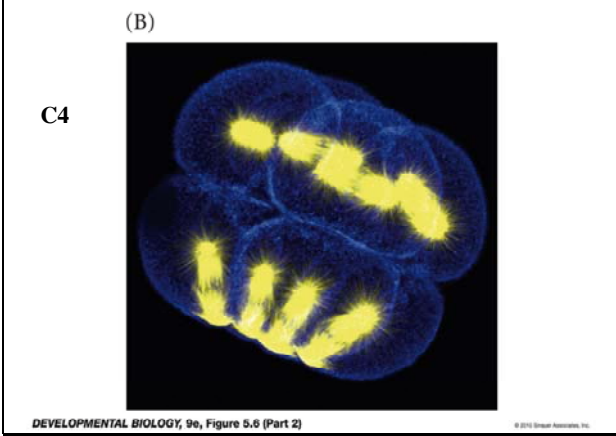
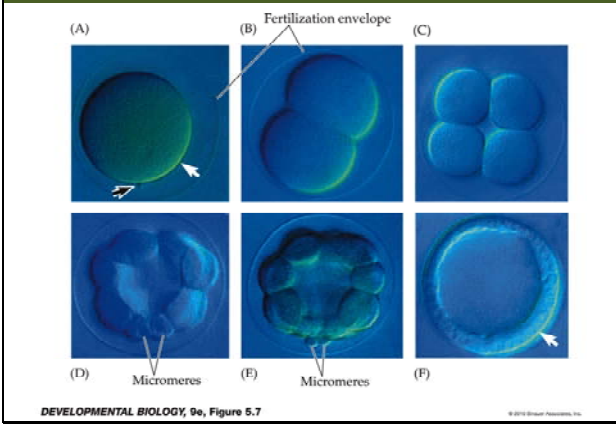


Figure 5.7 Micrographs of cleavage in live embryos of the sea urchin *Lytechinus variegatus*, seen from the side



- Proteins secreted from the inner surface of cells draw water from outside
- Results in hollow blastula

Figure 5.8 Fate map and cell lineage of the sea urchin *Strongylocentrotus purpuratus* (Part 1)

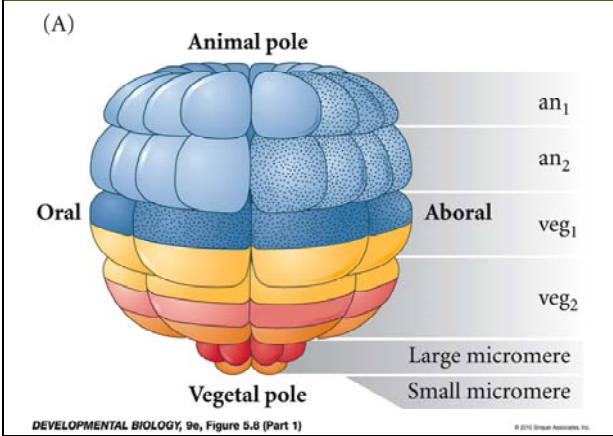
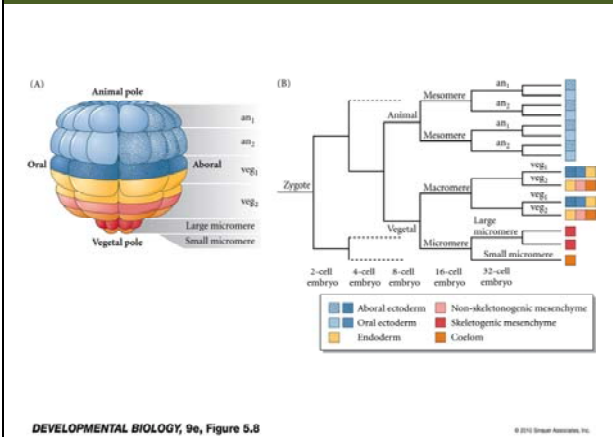


Figure 5.8 Fate map and cell lineage of the sea urchin *Strongylocentrotus purpuratus*



Micromeres induce presumptive ectodermal cells to acquire other fates

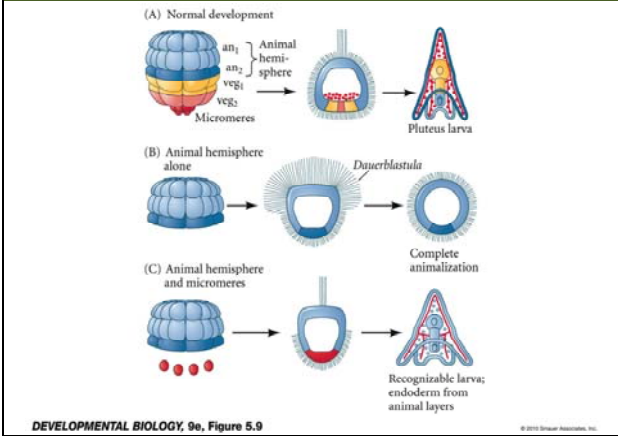
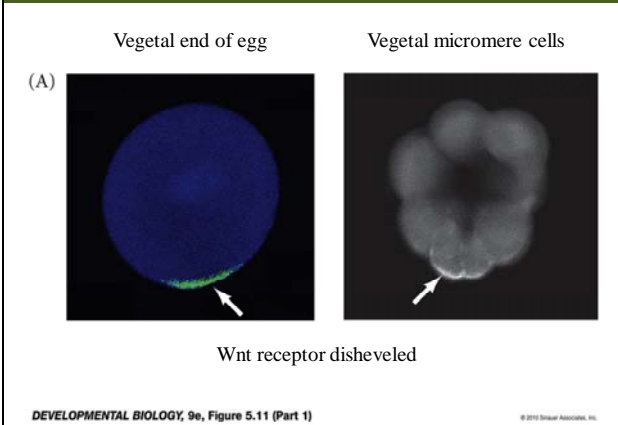


Figure 5.11 Role of Disheveled and β -catenin proteins in specifying the vegetal cells of the sea urchin embryo (Part 1)



β -catenin does the Wnt job

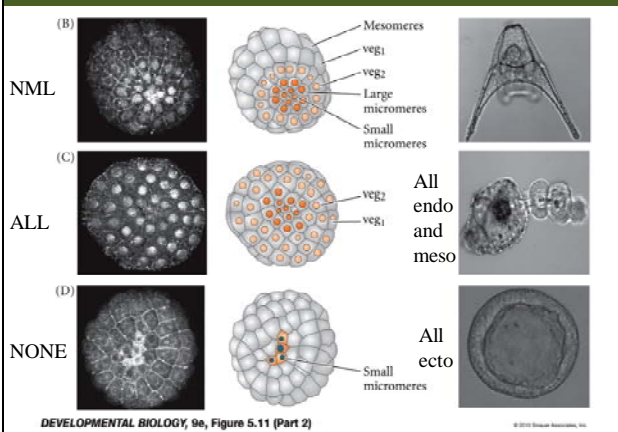
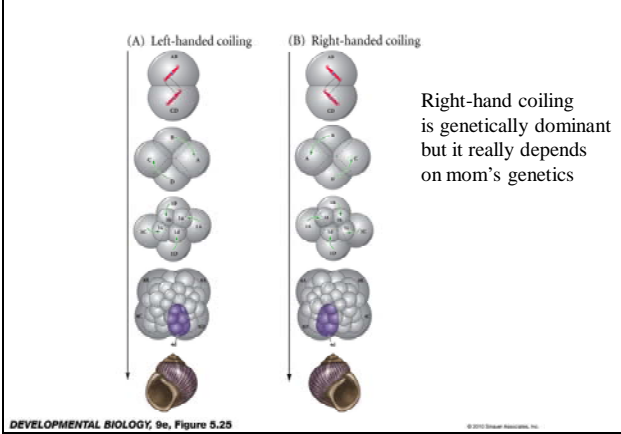
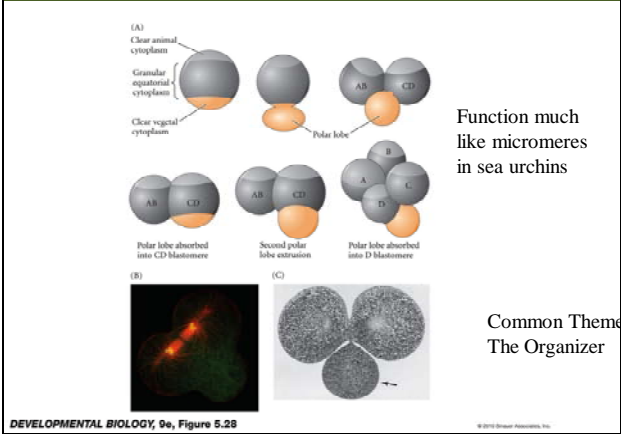


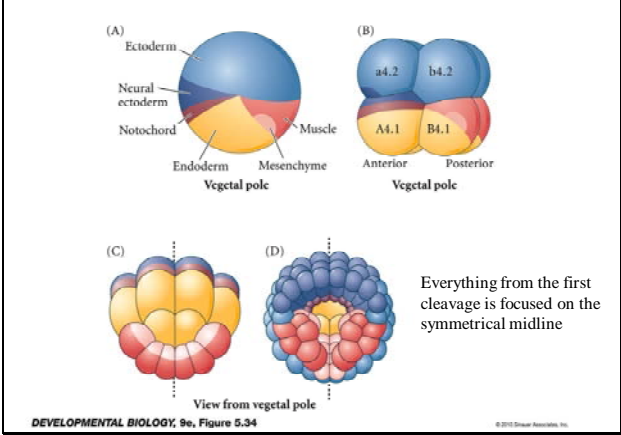
Figure 5.25 Looking down on the animal pole of left-coiling (A) and right-coiling (B) snails

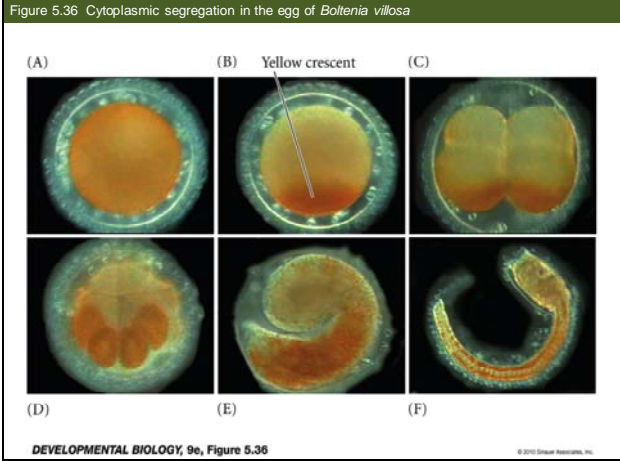


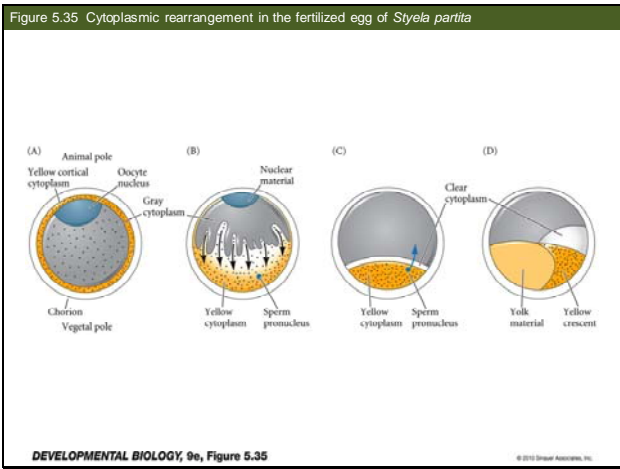
Polar lobe formation in certain mollusc embryos

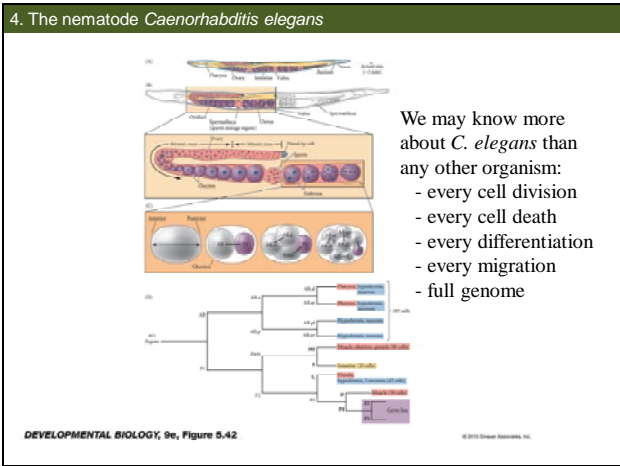


3. Bilateral, Holoblastic Cleavage in Tunicates

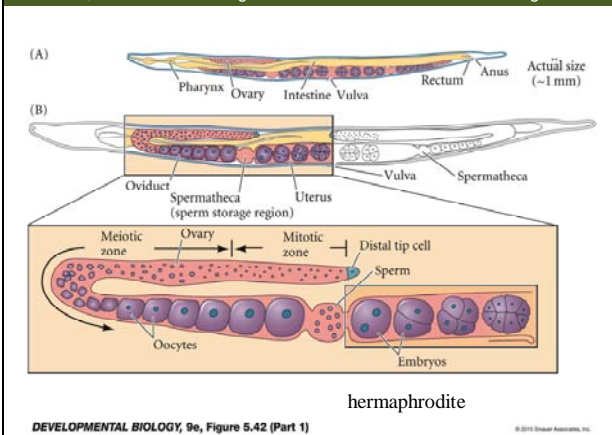








Rotational, Holoblastic Cleavage in the nematode *Caenorhabditis elegans*

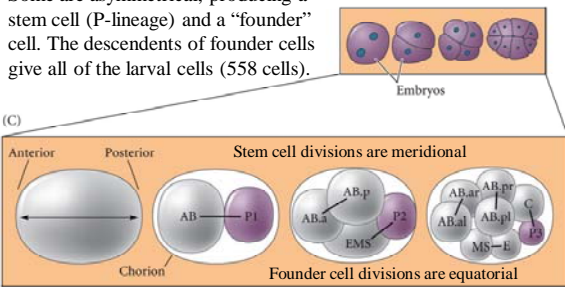


DEVELOPMENTAL BIOLOGY, 9e, Figure 5.42 (Part 1)

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Figure 5.42 The nematode *Caenorhabditis elegans* (Part 2)

Cleavage divisions drive rotation:
Some are asymmetrical, producing a stem cell (P-lineage) and a “founder” cell. The descendents of founder cells give all of the larval cells (558 cells).



An adult hermaphrodite has 959 cells, males have 1031

DEVELOPMENTAL BIOLOGY, 9e, Figure 5.42 (Part 2)

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5. Superficial cleavage in a *Drosophila* embryo



DEVELOPMENTAL BIOLOGY, 9e, Figure 6.1

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Figure 6.2 Nuclear and cell division in *Drosophila*

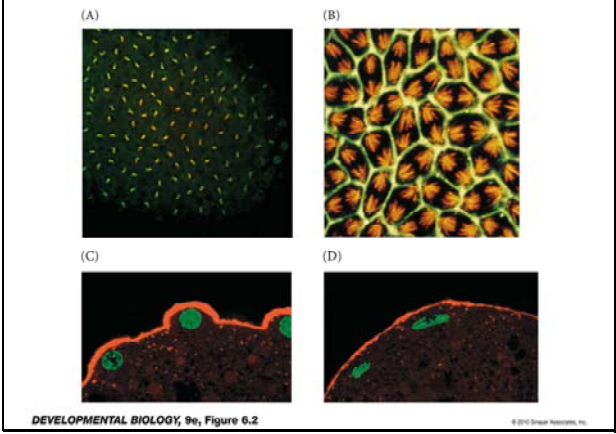
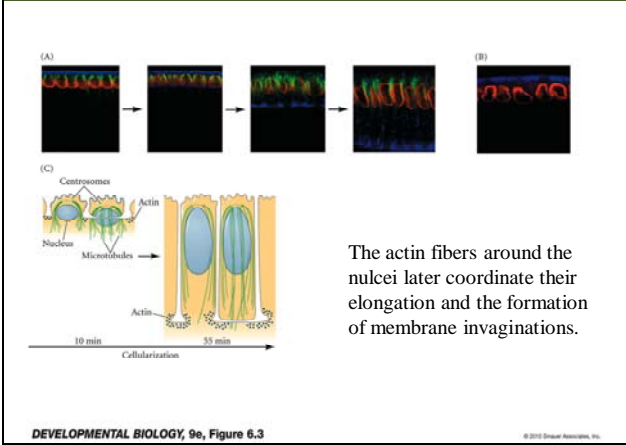


Figure 6.3 Formation of the cellular blastoderm in *Drosophila*

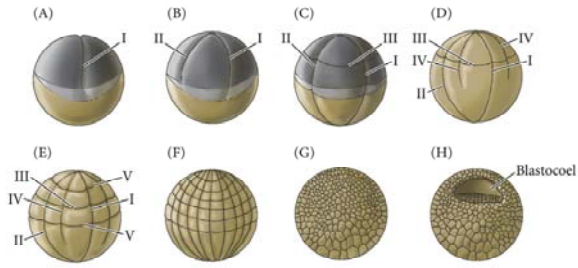


C. Vertebrates

1. The Frog
2. Zebrafish
3. The Chick
4. Human

Radial, Holoblastic Cleavage of a frog egg

Same basic design as the sea urchin

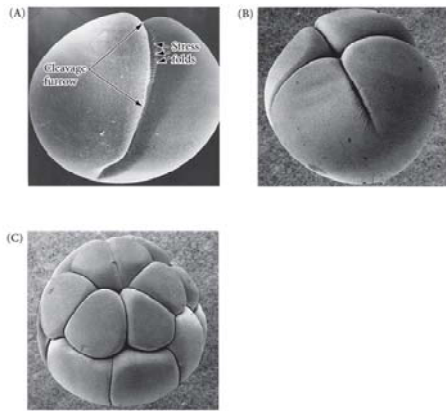


It is unequal because of the large amount of yolk in the vegetal end

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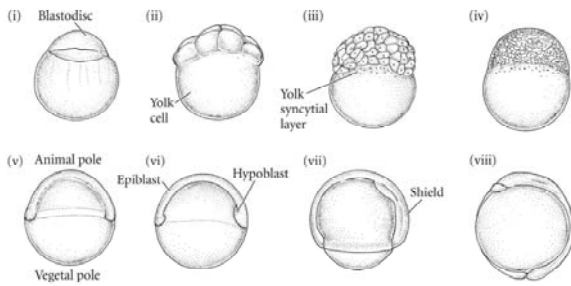
Figure 7.3 Scanning electron micrographs of frog egg cleavage



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2. Discoidal, Meroblastic Cleavage in Zebrafish



Very high yolk content

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Figure 7.40 Discoidal meroblastic cleavage in a zebrafish egg

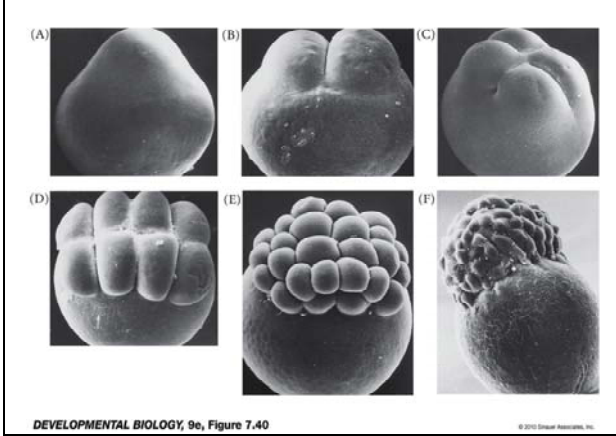
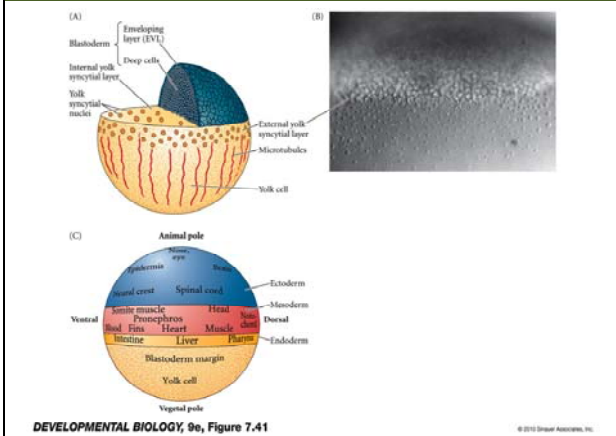


Figure 7.41 Fish blastula



3. Discoidal, Meroblastic Cleavage in a chick egg

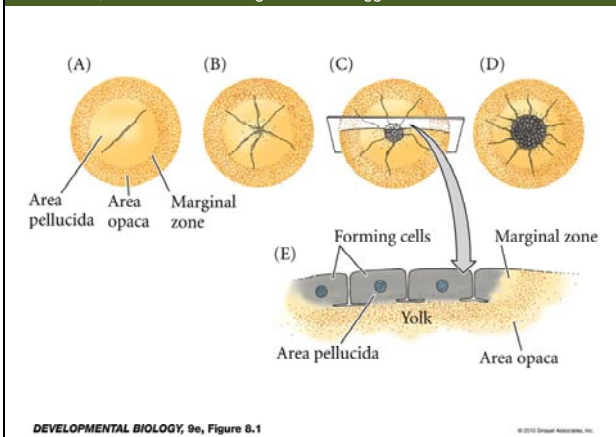
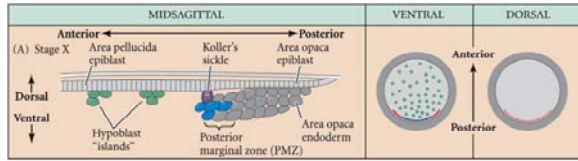


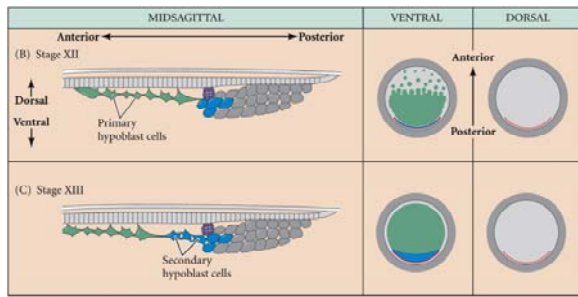
Figure 8.2. Formation of the chick blastoderm (Part 1)



DEVELOPMENTAL BIOLOGY, 9e, Figure 8.2 (Part 1)

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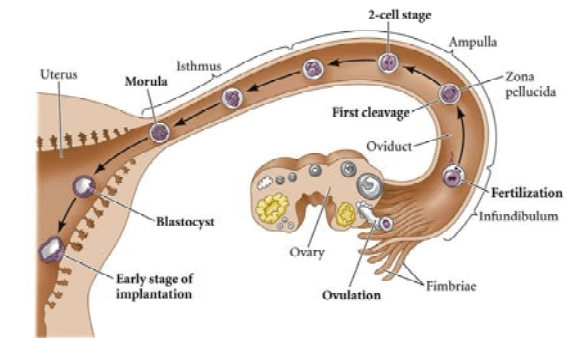
Figure 8.2. Formation of the chick blastoderm (Part 2)



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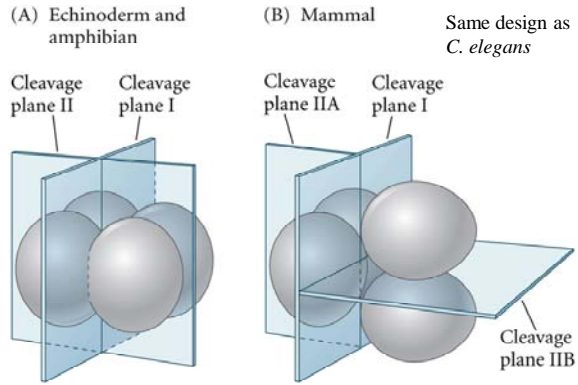
4. Mammals: Development of a human embryo from fertilization to implantation



DEVELOPMENTAL BIOLOGY, 9e, Figure 8.15

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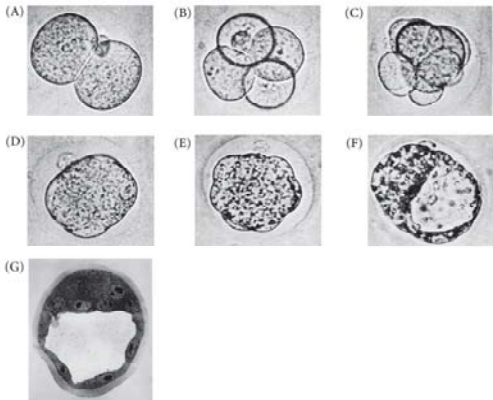
Rotational, Holoblastic Cleavage in Mammals



DEVELOPMENTAL BIOLOGY, 9e, Figure 8.16

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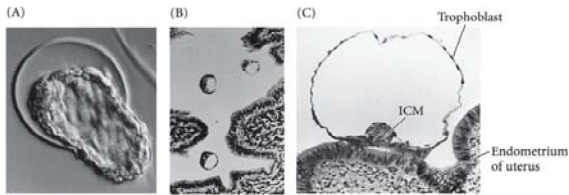
Figure 8.17 Cleavage of a single mouse embryo in vitro



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Figure 8.20 Hatching from the zona and implantation of the mammalian blastocyst in the uterus



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